Aminosubstituted α -D-glucosylmethylbenzenes (benzyl α -C-glucosides) and an N-(C- α -D-glucosylmethyl)aniline (anilinomethyl α -C-glucoside); novel α -D-glucosidase inhibitors

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ABSTRACT

D-Glucose was transformed via 3,4,5,7-tetra-O-benzyl-1,2-dideoxy-D-gluco-hept-1-enitol (5) into 2,6anhydro-3,4,5,7-tetra-O-benzyl-1-C-phenyl-p-erythro-L-ido-heptitol (9) the structure of which was determined by X-ray analysis of the per-O-acetylated derivative 12. 1-O-Mesylation of 9 and azide displacement gave only low yields of 2,6-anhydro-7-azido-1,3,4,5-tetra-O-benzyl-7-deoxy-7-C-phenyl-D-erythro-Lgulo-heptitol (16). Therefore, 9 was oxidized to 2,6-anhydro-3,4,5,7-tetra-O-benzyl-1-C-phenyl-D-glycerop-ido-heptose (15) and thence transformed into the (E/Z)-oximes 17h, which, with LiAlH₄ as reducing agent, gave 7-amino-2,6-anhydro-1,3,4,5-tetra-O-benzyl-7-deoxy-7-C-phenyl-D-erythro-L-guloheptitol (19), 7-amino-2,6-anhydro-1,3,4,5-tetra-O-benzyl-7-deoxy-7-C-phenyl-L-threo-L-gulo-heptitol (23), and 2,6-anhydro-1,3,4,5-tetra-O-benzyl-7-deoxy-7-phenylamino-p-glycero-L-gulo-heptitol (27) in 1:1:1 ratios. Their N-protection, hydrogenolytic O-debenzylation, and N-deprotection afforded the desired target molecules 7-amino-2,6-anhydro-7-deoxy-7-C-phenyl-p-erythro-L-gulo-heptitol (1a), 7amino-2,6-anhydro-7-deoxy-7-C-phenyl-L-threo-L-gulo-heptitol (2a), and 2,6-anhydro-7-deoxy-7-phenylamino-p-glycero-L-gulo-heptitol (4a). Hydrogenolysis of 15 furnished directly 2,6-anhydro-7-deoxy-7-Cphenyl-D-glycero-L-gulo-heptitol (3a). Inhibition studies with α -D-glucosidase from yeast with pnitrophenyl α -D-glucopyranoside as substrate exhibited, for 1a and 4a, K_i values of the same order as found for 1-deoxynojirimycin.

INTRODUCTION

The finding that amino sugar-based glycosidase inhibitors have potential as anti-HIV, diabetes, and cancer therapeutic agents has led to a wide interest and a demand for such compounds¹⁻⁶. Because phenyl glycosides are generally accepted as substrates by glycosidases, we planned to investigate novel aminosubstituted α -and β -D-glucopyranosylmethylbenzenes of the structure 1a, b and 2a, b (Scheme 1) as potential glucosidase inhibitors. These diastereomers should also provide infor-

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Scheme 1.

mation about the mechanism of action and the topography of the active site of the enzyme⁷⁻¹⁰. Their inhibitory action can be compared with that of the corresponding C-glucosyl compounds 3a,b, and the anilino derivatives 4a,b. Aniline itself¹¹ and aniline N-substituted with a galactopyranosylmethylene group¹² have already been shown to be inhibitors of glycosidases. In a previous publication⁹, we described syntheses and kinetic properties of the β anomers 1b-3b. In this paper, we report our results with the α anomers 1a-4a.

RESULTS AND DISCUSSION

Synthesis.—For the synthesis of the target molecules 1a-4a, the hydroxy-substituted α -D-glucosylmethylbenzene 9 was envisaged as starting material. To this end, the readily available 2,3,4,6-tetra-O-benzyl-D-glucose¹³ was transformed by a Wittig reaction into the known open-chain methylene derivative $5^{14,15}$ (Scheme 2). Subsequent ring closure by oxymercuration (\rightarrow 6) and oxidative demercuration following known procedures^{14,16} afforded exclusively C- α -D-glucopyranosylmethanol (7)^{14,17}. Oxidation under Swern conditions¹⁸ furnished the configurationally labile aldehyde 8 which, without isolation, was immediately treated with phenylmagnesium bromide in THF to furnish the benzylic alcohol 9 in good yield exclusively as the (1S)-diastereomer. Compound 9 was characterized as its O-acetyl derivative 10. The configuration of the newly formed stereocenter at C-1 was assigned by X-ray structure analysis of the per-O-acetylated derivative 12 (Fig. 1). This compound was obtained by hydrogenolytic O-debenzylation of 9 with palla-

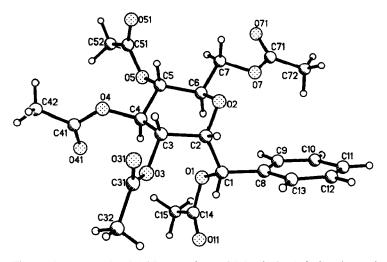


Fig. 1. Structure of 1,3,4,5,7-penta-O-acetyl-2,6-anhydro-1-C-phenyl-D-erythro-L-ido-heptitol (12) as found in the crystal.

dium-on-carbon as catalyst, furnishing the fully O-deprotected compound 11, followed by O-acetylation with acetic anhydride in pyridine.

The X-ray structure analysis of 12 exhibits slight chair deformation in order to avoid unfavourable 1,3-diaxial interactions of the axial α -hydroxybenzyl group at C-2 with the axial hydrogen atoms at C-4 and C-6, thus leading to a half-chair-like conformation. Comparison of the ¹H NMR coupling constants of pyranose ring protons shows that tertiary groups at the anomeric centre of C-glycosyl compounds generally tend to switch over from an axial into an equatorial position¹⁹. Contrastingly, substitution of the glycosidic oxygen by a methylene group induces practically

D-Gic
$$\frac{OR}{RO}$$
 $\frac{OR}{RO}$ $\frac{OR}{RO}$

Scheme 2.

no ring deformation as found for the C- α -D-glucopyranosylmethylbenzene 18 (see below) and in previous studies 20,21 .

For the introduction of the amino group, phenylcarbinol derivative 9 was activated by 1-O-mesylation to afford 13 (Scheme 3). Under standard reaction conditions (methanesulfonyl chloride-pyridine, one equivalent each in CH_2Cl_2), no reaction took place. Replacement of the solvent by pyridine gave exclusively the known elimination product 14^{17} . Best results for the synthesis of 13 were obtained with 4-dimethylaminopyridine and a twentyfold excess of methanesulfonyl chloride in CCl_4 . Treatment of 13 with tetramethylguanidinium azide in DMF afforded azido compound 16 together with 14 as major products. Other reaction conditions for the synthesis of 16 did not lead to improved results. For analytical purposes, the azido group was reduced with lithium aluminium hydride (LiAlH₄) in THF to give the $C-\alpha$ -D-glucopyranosylbenzylamine 19.

Because of low yields and accessibility of only one diastereoisomer, a second approach for the synthesis of compounds 1a and 2a was investigated. Swern oxidation of 9 yielded heptose derivative 15 in high yield. Treatment of 15 with hydroxylammonium chloride in pyridine furnished quantitatively the diastereomeric oximes 17h,l in a 1:1 ratio. The distinction h/l (high and low) refers to the difference in their R_f values; the absolute configuration of the (E/Z)-oximes 17h,l was not established since a mixture was sufficient in the following step. Complex hydrides are known to reduce oximes to amines with partial rearrangement to the corresponding N,N-disubstituted amine (via Beckmann rearrangement and reduction)²²; thus, not only access to the target molecules 1a and 2a but also to 4a was expected via such a route. Indeed, reduction of 17h,l with LiAlH₄ in THF afforded the diastereomeric amines 19 and 23 together with the rearranged anilino derivative 27 ($\sim 1:1:1$ ratios) in good overall yield.

According to our experience, hydrogenolytic O-debenzylation of sugars in the presence of unprotected amino groups often causes difficulties and ends up in partial debenzylation²³. Therefore, the following reaction sequence was chosen: 19 was N-protected by the *tert*-butoxycarbonyl (Boc) group under standard conditions, affording urethane 20; its hydrogenolytic O-debenzylation with palladium-on-carbon gave pure 21 which was isolated as its per-O-acetylated derivative 22. Acid-catalyzed cleavage of the Boc protecting group in 21 furnished the target molecule which was isolated as the trifluoroacetate 31 in quantitative yield. Its treatment with basic ion-exchange resin provided the aminosubstituted α -D-gluco-pyranosylmethylbenzene 1a. The same reaction sequence was applied, starting from 23, to obtain the Boc-protected urethane 24, the O-debenzylated intermediate 25, and its per-O-acetylated derivative 26. Cleavage of the Boc protecting group in 25 and then release of the amino group, as described above, afforded the diastereoisomers 32 and 2a, respectively, in quantitative yields.

In the case of the N-(C-glucosylmethyl)aniline 27, we took advantage of the trifluoroacetyl (TFA) protecting group, because Boc-protected secondary amines sometimes underwent side reactions during acid cleavage²³. Therefore, 27 was

treated with trifluoroacetic anhydride and pyridine in CH_2Cl_2 to give 28 in high yield. Hydrogenolysis under standard conditions afforded the O-debenzylated intermediate 29 which was transformed into 30 upon O-acetylation. Treatment of

TFA-protected 29 with sodium carbonate in aqueous methanol furnished the target molecule, which was isolated as the trifluoroacetate 33. The amino group was again released by treatment with basic ion-exchange resin to yield the N-(C-glucosylmethyl)aniline 4a.

The desired α -D-glucosylmethylbenzene 3a was readily obtained by direct hydrogenolytic O-debenzylation of 15 with palladium-on-carbon as catalyst which results also, as previously observed for the β anomer⁹, in removal of the oxygen next to the phenyl group. O-Acetylation of the crude product with acetic anhydride in pyridine furnished the deoxy compound 18, which was treated with methanolic sodium methoxide to afford compound 3a.

The ¹H NMR data of the intermediates and of the final products are in accordance with the assigned structures. All target molecules are stable in aqueous solution.

Inhibition studies.—The assay method was based on measuring the continuous release of p-nitrophenol from p-nitrophenyl α -D-glucopyranoside (PNPG) by the action of yeast α -D-glucosidase (maltase)²⁴. The Michaelis—Menten constant ($K_{\rm M}$) at pH 6.5 was determined to be $2 \cdot 10^{-4}$ M. For practical reasons, instead of the free amines 1a, 2a, and 4a, their trifluoroacetic acid salts 31–33 were used. The kinetic parameters were determined by Lineweaver—Burk plots²⁵. Table I summarizes the resulting inhibition constants ($K_{\rm i}$) together with those of some important known inhibitors. As expected, the α -D-glucosylmethylbenzene 3a showed only low inhibition and there is a clear distinction between the diastereomeric α -aminosubstituted derivatives 1a and 2a with a relatively strong inhibition for isomer 1a. However, it is remarkable that the N-(C-glucosylmethyl)aniline 4a yields a $K_{\rm i}$ value practically identical to that of 1-deoxynojirimycin. These observations give an indication of the relative position of acid groups in the active site of the enzyme which participate in the glycosidic cleavage.

EXPERIMENTAL

General methods.—Melting points are uncorrected. Optical rotations were measured at 22°C with a Perkin-Elmer 241 MC polarimeter. 1 H NMR and 13 C NMR spectra were recorded for solutions in CDCl₃ (internal standard Me₄Si) with a Bruker AC 250 Cryospec instrument. R_f values refer to TLC performed on Silica Gel 60 F₂₅₄ (Merck). Column chromatography was performed under normal pressure with silica gel (Merck, 70–230 mesh ASTM and 230–400 mesh ASTM for flash chromatography) and under elevated pressure with LiChroprep Si 60 (Merck, $15-25~\mu m$). The bp of the light petroleum was $35-65^{\circ}$ C. UV spectra were recorded with a Philips PU 8740 UV/VIS spectrophotometer. IR spectra were recorded with a Mattson Polaris FT-IR spectrometer.

2,6-Anhydro-3,4,5,7-tetra-O-benzyl-1-C-phenyl-D-erythro-L-ido-heptitol (9).—A solution of oxalyl chloride (370 μ L, 23.8 mmol) in dry CH₂Cl₂ (10 mL) was cooled to -60° C and a solution of dry Me₂SO (670 μ L, 51.9 mmol) in dry CH₂Cl₂ (2 mL)

TABLE I			
Type of inhibition	and	inhibition	constants $(K_i)^a$

Compound	pН	Inhibition type	$K_{i}[M]$	Ref
Maltose	6.8	(substrate)	$3.5 \cdot 10^{-2 \ b}$	24
3a	6.5	competitive	$1.3 \cdot 10^{-3}$	
2a	6.5	competitive	$1.1 \cdot 10^{-3}$	
Phenyl α-D-glucopyranoside	6.8	(substrate)	$4.0 \cdot 10^{-3 \ b}$	24
N-Methyldeoxynojirimycin	6.5	competitive	$3.7 \cdot 10^{-4}$	26
1a	6.5	competitive	$3.8 \cdot 10^{-5}$	
4a	6.5	competitive	$1.1 \cdot 10^{-5}$	
Deoxynojirimycin	6.5	competitive	$0.9 \cdot 10^{-5}$	26

^a For method, see Experimental. ^b With respect to PNPG as substrate.

was added dropwise during ca. 5 min. Stirring was continued at -60° C for 10 min, followed by addition of a solution of 2,6-anhydro-1,3,4,5-tetra-O-benzyl-D-glycero-L-gulo-heptitol¹⁴⁻¹⁶ (7; 2.18 g, 3.93 mmol) in dry CH₂Cl₂ (4 mL) during ca. 5 min. The mixture was stirred for 15 min followed by addition of Et₃N (2.7 mL, 19.6 mmol) during ca. 5 min. After 10 min, the cold mixture was diluted with CH₂Cl₂ (50 mL) and poured into aq NH₄Cl (50 mL), and the organic layer was collected. The aqueous phase was re-extracted with CH₂Cl₂ (20 mL). The combined organic layers were dried (MgSO₄) and evaporated to dryness. The crude aldehyde was dissolved in dry diethyl ether (40 mL) and added dropwise to a Grignard solution (20 mL of a 0.3 M solution in diethyl ether) prepared from bromobenzene and magnesium. After warming at 35°C for 30 min, the mixture was cooled and then decomposed by adding, with stirring, aq 20% NH₄Cl (50 mL). The aqueous phase was re-extracted with diethyl ether (20 mL), and the combined organic layers were washed successively with aq NH₄Cl (40 mL) and brine (40 mL). The organic layer was dried (MgSO₄) and concentrated under reduced pressure. Flash chromatography (4:1 light petroleum-EtOAc) of the residue yielded 9 (1.98 g, 80%), isolated as a colorless oil; $[\alpha]_D + 39.5^\circ$ (c 1, CHCl₃), R_f 0.33 (3:1 light petroleum-EtOAc); ¹H NMR (250 MHz, CDCl₃): δ 2.55 (bs, 1 H, OH), 3.55 (dd, 1 H, J 4.7, 6.1 Hz, H-3), 3.64 (dd, 1 H, J 6.1, 8.2 Hz, H-5), 3.63–3.73 (m, 2 H, H-7a,7b), 3.99–4.05 (m, 2 H, H-2,4), 4.17 (d, 1 H, J 11.5 Hz, CH₂Ph), 4.18-4.27 (m, 1 H, H-6), 4.44-4.71 (m, 7 H, CH_2Ph), 5.08 (d, 1 H, $J_{1,2}$ 7.2 Hz, H-1), 7.15–7.36 (m, 25 H, 5 Ph). Anal. Calcd for C₄₁H₄₂O₆ · 0.5H₂O: C, 76.96; H, 6.77. Found: C, 76.97; H, 6.82.

1-O-Acetyl-2,6-anhydro-3,4,5,7-tetra-O-benzyl-1-C-phenyl-D-erythro-L-ido-heptitol (10).—To a solution of 9 (40 mg, 0.063 mmol) in dry pyridine (5 mL) was added Ac₂O (5 mL), and the mixture was left at room temperature overnight. When the reaction was complete as shown by TLC [R_f 0.37 (4:1 light petroleum-EtOAc)], the mixture was coevaporated with toluene (3 × 20 mL). Flash chromatography (6:1 light petroleum-EtOAc) of the residue gave 10 (42 mg, 98%) isolated as a colorless oil; [α]_D +59° (c 1, CHCl₃); ¹H NMR (250 MHz, CDCl₃): δ 2.04 (s, 3 H, OAc), 3.60-3.70 (m, 4 H, H-3,4,7a,7b), 3.99 (t, 1 H, J 7.2 Hz, H-4), 4.11-4.17 (m, 1

H, H-6), 4.22 (t, 1 H, J 6.4 Hz, H-2), 4.20–4.77 (m, 8 H, CH_2 Ph), 6.19 (d, 1 H, J 6.4 Hz, H-1), 7.17–7.31 (m, 25 H, 5 Ph). Anal. Calcd for $C_{43}H_{44}O_7$: C, 76.76; H, 6.59. Found: C, 76.37; H, 6.56.

2,6-Anhydro-1-C-phenyl-D-erythro-L-ido-heptitol (11).— To a solution of 12 (140 mg, 0.291 mmol) in dry MeOH (25 mL) was added sodium methoxide (0.2 mL of a 0.2 M solution in MeOH). After 5 h at room temperature, the mixture was neutralized with Amberlite IR-120 (H⁺) resin, filtered, and concentrated under reduced pressure. Flash chromatography (9:1 CHCl₃-MeOH) of the residue gave 11 (75 mg, 96%) as a colorless oil; $[\alpha]_D + 80^\circ$ (c 1, MeOH); R_f 0.08 (9:1 CHCl₃-MeOH); ¹H NMR (250 MHz, D₂O): δ 3.31 (t, 1 H, J 7.8 Hz, H-5), 3.40 (dd, 1 H, $J_{2,3}$ 5.1, $J_{3,4}$ 7.6 Hz, H-3), 3.55 (dd, 1 H, $J_{6,7a}$ 3.5, J_{gem} 12.5 Hz, H-7a), 3.62 (dd, 1 H, $J_{6,7b}$ 5.7, J_{gem} 12.5 Hz, H-7b), 3.83 (t, 1 H, J 7.6 Hz, H-4), 3.89-3.95 (m, 2 H, H-2,6), 4.93 (d, 1 H, J 6.4 Hz, H-1), 7.18-7.26 (m, 5 H, Ph).

1,3,4,5,7-Penta-O-acetyl-2,6-anhydro-1-C-phenyl-D-erythro-L-ido-heptitol (12).— To a solution of 9 (1.5 g, 2.38 mmol) in EtOAc (35 mL) and MeOH (35 mL) was added palladium-on-carbon (10%, 150 mg). After hydrogenolysis for 6 h, the mixture was filtered and concentrated under reduced pressure. The residue was dissolved in dry pyridine (20 mL) and treated with Ac₂O (20 mL), and the mixture was left at room temperature overnight. When the reaction was complete as shown by TLC $[R_f \ 0.5 \ (1:1 \ light \ petroleum-EtOAc)]$, the mixture was coevaporated with toluene $(3 \times 50 \text{ mL})$. Flash chromatography $(3:1 \rightarrow 2:1 \text{ light petroleum-EtOAc})$ of the residue yielded 12 (800 mg, 70%) as a colorless oil that crystallized on standing; $[\alpha]_D + 56^\circ$ (c 1, CHCl₃). Careful recrystallization from light petroleum– EtOAc gave crystals, mp 147°C, which were suitable for X-ray analysis; ¹H NMR (250 MHz, CDCl₃): δ 1.78 (s, 3 H, OAc), 1.99 (s, 6 H, 2 OAc), 2.02, 2.19 (2 s, 6 H, 2 OAc), 3.96 (dd, 1 H, $J_{6.7a}$ 2.4, J_{gem} 11.6 Hz, H-7a), 4.13 (dd, 1 H, $J_{6.7b}$ 5.8, J_{gem} 11.6 Hz, H-7b), 4.23 (ddd, 1 H, $J_{5,6}$ 8.4, $J_{6,7a}$ 2.4, $J_{6,7b}$ 5.8 Hz, H-6), 4.54 (t, 1 H, J6.4 Hz, H-2), 4.93–5.01 (m, 2 H, H-3,5), 5.58 (t, 1 H, J 8.7 Hz, H-4), 6.10 (d, 1 H, J 6.4 Hz, H-1), 7.24–7.34 (m, 5 H, Ph). Anal. Calcd for C₂₃H₂₈O₁₁: C, 57.50; H, 5.88. Found: C, 57.50; H, 5.90.

2,6-Anhydro-3,4,5,7-tetra-O-benzyl-1-O-methanesulfonyl-1-C-phenyl-D-erythro-Lido-heptitol (13).—To a solution of 9 (125 mg, 0.198 mmol) in dry CCl₄ (5 mL) were added 4-dimethylaminopyridine (50 mg) and methanesulfonyl chloride (200 μL). The mixture was stirred for 6 days at 50°C, then allowed to come to room temperature, diluted with diethyl ether (50 mL), and extracted with water (20 mL). The organic layer was washed with satd aq NaHCO₃ (20 mL) and brine (20 mL), and evaporated to dryness. Flash chromatography (19:1 toluene–EtOAc) yielded 13 (77 mg, 55%) as a colorless oil; $[\alpha]_D$ +58° (c 1, CHCl₃); R_f 0.45 (9:1 toluene–EtOAc); ¹H NMR (250 MHz, CDCl₃): δ 2.85 (s, 3 H, CH₃), 3.30 (t, 1 H, J 4.7 Hz, H-3), 3.61 (dd, 1 H, $J_{4,5}$ 5.0, $J_{5,6}$ 7.5 Hz, H-5), 3.65 (dd, 1 H, $J_{6,7a}$ 3.4, J_{gem} 10.4 Hz, H-7a), 3.73 (dd, 1 H, $J_{6,7b}$ 5.4, J_{gem} 10.4 Hz, H-7b), 3.87 (t, 1 H, J 5.0 Hz, H-4), 4.00 (d, 1 H, J 11.2 Hz, CH_2 Ph), 4.21–4.28 (m, 2 H, H-2,6), 4.36 (d, 1 H, J 11.2 Hz, CH_2 Ph), 4.44–4.63 (m, 6 H, CH_2 Ph), 5.80 (d, 1 H, J 7.5 Hz, H-1),

7.14–7.35 (m, 25 H, 5 Ph). Anal. Calcd for $C_{42}H_{44}O_8S$: C, 71.16; H, 6.22. Found: C, 71.05; H, 6.32.

2,6-Anhydro-3,4,5,7-tetra-O-benzyl-1-C-phenyl-D-glycero-D-ido-heptose (15).—A solution of oxalyl chloride (95 µL, 1.11 mL) in dry CH₂Cl₂ (5 mL) was cooled to -60° C and a solution of dry Me₂SO (170 μ L, 2.39 mmol) in dry CH₂Cl₂ (1 mL) was added dropwise during ca. 5 min. Stirring was continued at -60° C for 15 min followed by addition of a solution of 9 (450 mg, 0.713 mmol) in dry CH₂Cl₂ (2 mL) during ca. 10 min. The mixture was stirred for 30 min, then Et₃N (700 μ L, 5.05 mmol) was added dropwise during ca. 5 min and the solution was allowed to attain room temperature. The mixture was diluted with CH₂Cl₂ (60 mL), then extracted with aq NH₄Cl (4×20 mL), and the organic layer was concentrated in vacuo. Flash chromatography (5:1 light petroleum-EtOAc) yielded 15 (410 mg, 92%) as a colorless oil; $[\alpha]_D$ +22° (c 1, CHCl₃); R_f 0.48 (3:1 light petroleum-EtOAc); ¹H NMR (250 MHz, CDCl ₃): δ 3.57 (dd, 1 H, $J_{6,7a}$ 1.9, $J_{\rm gem}$ 10.7 Hz, H-7a), 3.70 (dd, 1 H, $J_{6.7b}$ 3.2, J_{gem} 10.7 Hz, H-7b), 3.75 (dd, 1 H, $J_{4.5}$ 8.3, $J_{5.6}$ 9.7 Hz, H-5), 3.84–3.90 (m, 1 H, H-6), 3.95 (dd, 1 H, $J_{2,3}$ 6.1, $J_{3,4}$ 8.9 Hz, H-3), 4.39–4.61 (m, 5 H, CH_2 Ph, H-4), 4.77-5.03 (m, 4 H, CH₂Ph), 5.22 (d, 1 H, J 6.1 Hz, H-2); 7.12-7.96 (m, 25 H, 5 Ph); 13 C NMR (62.5 MHz, CDCl₃): δ 196.95 (PhCO). Anal. Calcd for $C_{41}H_{40}O_6$: C, 78.32; H, 6.41. Found: C, 78.16; H, 6.43.

2,6-Anhydro-7-azido-1,3,4,5-tetra-O-benzyl-7-deoxy-7-C-phenyl-D-erythro-L-gulo-heptitol (16).—To a solution of 13 (40 mg, 0.056 mmol) in dry DMF (3 mL) was added tetramethylguanidinium azide (89 mg, 0.56 mmol). The mixture was stirred at 70°C for 7 days, then diluted with water (50 mL), and extracted with diethyl ether (2 × 30 mL). The combined organic layers were washed with water (4 × 15 mL) and concentrated under reduced pressure. Flash chromatography (50:1 toluene–EtOAc) of the residue yielded 16 (7.5 mg, 20%) as a colorless oil; $[\alpha]_D$ –21° (c 0.5, CHCl₃); R_f 0.44 (19:1 toluene–EtOAc); $\nu_{\rm max}^{\rm CCl_4}$ 2106.9 cm⁻¹ (N₃); ¹H NMR (250 MHz, CDCl₃): δ 3.36 (dd, 1 H, $J_{\rm gem}$ 10.7, $J_{\rm 1a,2}$ 3.8 Hz, H-1a), 3.45 (dd, 1 H, $J_{\rm gem}$ 10.7, $J_{\rm 1b,2}$ 4.6 Hz, H-1b), 3.63 (dd, 1 H, $J_{\rm 4.8}$, $J_{\rm 7.4}$ Hz, H-3); 3.81–3.92 (m, 3 H, H-2,4,5), 4.07 (dd, 1 H, $J_{\rm 5,6}$ 3.0, $J_{\rm 6,7}$ 9.6 Hz, H-6), 4.14–4.72 (m, 8 H, C H_2 Ph), 4.82 (d, 1 H, $J_{\rm 6,7}$ 9.6 Hz, H-7), 7.06–7.38 (m, 25 H, 5 Ph).

(E/Z)-2,6-Anhydro-3,4,5,7-tetra-O-benzyl-1-C-phenyl-D-glycero-D-ido-heptose oxime (17h,l).—To a solution of 15 (7.67 g, 11.9 mmol) in dry pyridine (100 mL) was added hydroxylammonium chloride (2.48 g, 35.7 mmol). After warming for 5 h at 40°C, the mixture was concentrated under reduced pressure, diluted with toluene (50 mL), and evaporated to dryness. Filtration over silica gel (9:1 toluene-acetone) yielded 17h,l (7.7 g, quant; E: Z = 1:1) as a colorless oil, which was used as the E/Z-mixture in the next step. Separation of the E/Z isomers was possible by flash chromatography (83:16:1 \rightarrow 74:25:1 light petroleum-EtOAc-Et₃N).

Compound 17h: R_f 0.26 (83:16:1 light petroleum–EtOAc–Et $_3$ N); $[\alpha]_D$ +26° (c 1, CHCl $_3$); 1H NMR (250 MHz, CDCl $_3$): δ 3.49 (dd, 1 H, $J_{\rm gem}$ 10.5, $J_{6,7a}$ 1.8 Hz, H-7a), 3.60 (dd, 1 H, $J_{\rm gem}$ 10.5, $J_{6,7b}$ 4.0 Hz, H-7b), 3.64 (dd, 1 H, J 8.8, 9.8 Hz, H-5), 3.86–3.91 (m, 1 H, H-6), 3.90 (dd, 1 H, $J_{2,3}$ 6.4, $J_{3,4}$ 9.5 Hz, H-3), 4.27–5.12

(m, 10 H, CH_2 Ph, H-2,4), 4.72 (d, 1 H, J 6.4 Hz, H-2), 7.08–7.46 (m, 25 H, 5 Ph), 9.22 (bs, 1 H, NOH); ¹³C NMR (62.5 MHz, CDCl₃): δ 152.8 (CNOH). Anal. (for 17h) Calcd for $C_{41}H_{41}NO_6$: C, 76.49; H, 6.42; N, 2.18. Found: C, 76.74; H, 6.53; N, 2.20.

Compound 17l: R_f 0.19 (83:16:1 light petroleum–EtOAc–Et₃N); $[\alpha]_D$ +7.0° (c 1, CHCl₃); ¹H NMR (250 MHz, CDCl₃): δ 3.48 (dd, 1 H, $J_{\rm gem}$ 10.7, $J_{6,7a}$ 2.9 Hz, H-7a), 3.54 (dd, 1 H, $J_{\rm gem}$ 10.7, $J_{6,7b}$ 3.6 Hz, H-7b), 3.84–3.94 (m, 3 H, H-4,5,6), 4.32–4.63 (m, 9 H, C H_2 Ph, H-3), 5.69 (d, 1 H, J 3.6 Hz, H-2), 7.18–7.37 (m, 23 H, Ph), 7.67–7.71 (m, 2 H, Ph), 9.26 (bs, 1 H, NOH); ¹³C NMR (62.5 MHz, CDCl₃): δ 161.1 (CNOH). Anal. (for 17l) Calcd for C₄₁H₄₁NO₆: C, 76.49; H, 6.42; N, 2.18. Found: C, 76.43; H, 6.50; N, 2.23.

1,3,4,5-Tetra-O-acetyl-2,6-anhydro-7-deoxy-7-C-phenyl-D-glycero-L-gulo-heptitol (18).—To a solution of 15 (170 mg, 0.27 mmol) in EtOAc (5 mL) and MeOH (5 mL) was added palladium-on-carbon (10%, 70 mg). After hydrogenolysis for 3 h, the mixture was filtered and concentrated under reduced pressure. A mixture of the residue, dry pyridine (5 mL), and Ac₂O (5 mL) was left for 3 h at room temperature. Then, the mixture was coevaporated with toluene (3 × 25 mL). Flash chromatography (3:1 light petroleum–EtOAc) of the residue yielded 18 (80 mg, 70%) as white crystals; mp 79°C; [α]_D +85° (c 1, Et₂O); R_f 0.64 (1:1 light petroleum–EtOAc); ¹H NMR (250 MHz, CDCl₃): δ 2.00, 2.01, 2.07, 2.09 (4 s, 12 H, 4 OAc), 2.94 (dd, 1 H, $J_{6,7a}$ 5, J_{gem} 14.9 Hz, H-7a), 3.10 (dd, 1 H, $J_{6,7b}$ 9.9, J_{gem} 14.9 Hz, H-7b), 4.02–4.10 (m, 2 H, H-1a,1b), 4.18–4.26 (m, 1 H, H-2), 4.55 (ddd, 1 H, $J_{5,6}$ = $J_{6,7a}$ = 5, $J_{6,7b}$ 9.9 Hz, H-6), 5.01 (t, 1 H, J 9 Hz, H-3), 5.15 (dd, 1 H, $J_{4,5}$ 9, $J_{5,6}$ 5 Hz, H-5), 5.47 (t, 1 H, J 9 Hz, H-4), 7.21–7.36 (m, 5 H, Ph). Anal. Calcd for C₂₁H₂₆O₉: C, 59.71; H, 6.20. Found: C, 59.72; H, 6.28.

7-Amino-2,6-anhydro-1,3,4,5-tetra-O-benzyl-7-deoxy-7-C-phenyl-p-erythro-L-guloheptitol (19), 7-amino-2,6-anhydro-1,3,4,5-tetra-O-benzyl-7-deoxy-7-C-phenyl-Lthreo-L-gulo-heptitol (23), and 2,6-anhydro-1,3,4,5-tetra-O-benzyl-7-deoxy-7-phenylamino-D-glycero-L-gulo-heptitol (27).—Procedure (a). To a suspension of LiAlH (350 mg) in dry THF (110 mL) was added dropwise a solution of 17h,l (3.7 g, 5.75 mmol) in dry THF (40 mL) at room temperature. The mixture was heated under reflux (bath temperature, 75°C) for 60 min, then allowed to attain room temperature, and filtered. The filtrate was treated successively with wet diethyl ether (10 mL) and water (3 mL). The mixture was again filtered, and the filtrate was washed with aq NH₄Cl (100 mL). The aqueous phase was re-extracted with diethyl ether (100 mL), and the combined organic layers were concentrated under reduced pressure and coevaporated with toluene (2 × 50 mL). Flash chromatography $(100:2 \rightarrow 95:5 \rightarrow 9:1 \text{ CHCl}_3\text{-MeOH})$ yielded 19 (1.04 g, 29%) as a yellowish oil that crystallized on standing, 23 (880 mg, 24%) as a colorless oil, and crude 27 (900 mg). Compound 27 was purified further by flash chromatography (9:1 light petroleum-EtOAc), followed by MPLC (4:1 light petroleum-EtOAc), to give pure 27 (725 mg, 20%) as a colorless oil. Compound 19 could be recrystallized

from light petroleum-EtOAc and from hot EtOH to form colorless needles; mp 113°C.

Procedure (b) for 19. To a suspension of LiAlH₄ (5 mg) in dry THF (1 mL) was added dropwise a solution of 16 (7.5 mg, 11.6 μ mol) in dry THF (1 mL) with stirring. The mixture was heated under reflux for 90 min, then allowed to attain room temperature, and successively treated with wet diethyl ether (5 mL) and water (1 drop). The solution was filtered (Celite), the filter cake was washed with diethyl ether (10 mL), and the combined filtrates were evaporated to dryness. Flash chromatography (74:25:1 light petroleum-EtOAc-Et₃N) yielded 19 (4 mg, 56%) as a colorless oil.

Compound 19: R_f 0.34 (100:2 CHCl₃–MeOH); $[\alpha]_D$ +22.5° (c 1, CHCl₃); 1H NMR (250 MHz, CDCl₃): δ 1.74 (bs, 2 H, NH₂), 3.21 (dd, 1 H, $J_{\rm gem}$ 10.7, $J_{\rm 1a,2}$ 2.2 Hz, H-1a), 3.46 (dd, 1 H, $J_{\rm gem}$ 10.7, $J_{\rm 1b,2}$ 3.4 Hz, H-1b), 3.60–3.71 (m, 2 H, H-2,3), 3.93 (dd, 1 H, $J_{\rm 4,5}$ 7, $J_{\rm 5,6}$ 4.4 Hz, H-5), 4.00 (t, 1 H, J 7 Hz, H-4), 4.14–4.88 (m, 10 H, 4 C H_2 Ph, H-6,7), 4.20 (dd, 1 H, $J_{\rm 5,6}$ 4.4, $J_{\rm 6,7}$ 9.2 Hz, H-6), 4.40 (d, 1 H, J 9.2 Hz, H-7), 7.12–7.40 (m, 25 H, 5 Ph). Anal. (for 19) Calcd for C₄₁H₄₃NO₅: C, 78.19; H, 6.88; N, 2.22. Found: C, 77.94; H, 6.95; N, 2.42.

Compound **23**: R_f 0.22 (100:2 CHCl₃-MeOH); $[\alpha]_D$ +28° (c 1, CHCl₃); 1H NMR (250 MHz, CDCl₃): δ 1.84 (bs, 2 H, NH₂), 3.32 (dd, 1 H, $J_{4,5}$ 5.3, $J_{5,6}$ 3.6 Hz, H-5), 3.63 (dd, 1 H, $J_{2,3}$ 7.4, $J_{3,4}$ 5.3 Hz, H-3), 3.71–3.73 (m, 2 H, H-1a,1b), 3.82 (t, 1 H, J 5.3 Hz, H-4), 3.92 (d, 1 H, J 11.3 Hz, CHHPh), 4.03 (dd, 1 H, $J_{5,6}$ 3.6, $J_{6,7}$ 9.2 Hz, H-6), 4.08–4.15 (m, 1 H, H-2), 4.25 (d, 1 H, J 11.3 Hz, CHJ Ph), 4.35 (d, 1 H, J 9.2 Hz, H-7), 4.47–4.65 (m, 6 H, 3 CJ Ph), 7.04–7.32 (m, 25 H, 5 Ph). Anal. (for **23**) Calcd for CJ Calcd for C

Compound 27: R_f 0.86 (100:2 CHCl₃–MeOH); $[\alpha]_D$ +44.5° (c 1, CHCl₃); 1H NMR (250 MHz, CDCl₃): δ 3.40 (dd, 1 H, $J_{6,7a}$ 10.9, J_{gem} 12.7 Hz, H-7a), 3.48 (dd, 1 H, $J_{6,7b}$ 4.9, J_{gem} 12.7 Hz, H-7b), 3.64–3.92 (m, 6 H, H-1a, 1b, 2, 3, 4, 5), 4.31–4.39 (m, 1 H, H-6), 4.49–5.00 (m, 8 H, 4 C H_2 Ph), 6.59–6.62 (m, 2 H, Ph), 6.71–6.76 (m, 1 H, Ph), 7.14–7.38 (m, 22 H, Ph). Anal. (for 27) Calcd for C $_{41}H_{43}$ NO₅: C, 78.19; H, 6.88; N, 2.22. Found: C, 78.21; H, 6.95; N, 2.17.

2,6-Anhydro-1,3,4,5-tetra-O-benzyl-7-tert-butoxycarbonylamino-7-deoxy-7-C-phenyl-D-erythro-L-gulo-heptitol (20).—To a solution of 19 (125 mg, 0.198 mmol) in dioxane (10 mL) was added (t-BuOCO)₂O (160 mg, 0.733 mmol) and Et₃N (1 mL). The mixture was left for 4 h at room temperature, then concentrated under reduced pressure and coevaporated with toluene (25 mL). Flash chromatography (7:1 → 5:1 light petroleum–EtOAc) yielded 20 (100 mg, 69%) as a colorless glassy solid; mp 109°C; $[\alpha]_D$ +21° (c 1, CHCl₃); R_f 0.41 (4:1 light petroleum–EtOAc). H NMR (250 MHz, CDCl₃): δ 1.37 [s, 9 H, C(CH₃)₃], 3.53–3.68 (m, 3 H, H-1a, 1b,5), 3.57 (dd, 1 H, J_{gem} 10.7, $J_{1a,2}$ 3.1 Hz, H-1a), 3.65 (dd, 1 H, J_{gem} 10.7, $J_{1b,2}$ 3.8 Hz, H-1b), 3.74 (dd, 1 H, $J_{2,3}$ 8.2, $J_{3,4}$ 6 Hz, H-3), 3.91 (t, 1 H, J 6 Hz, H-4), 3.93–4.00 (m, 1 H, H-2), 4.15 (dd, 1 H, $J_{5,6}$ 3.5, $J_{6,7}$ 6.7 Hz, H-6), 4.36–4.70 (m, 8 H, 4 C H_2 Ph), 5.14 (bt, 1 H, J 7 Hz, H-7), 5.93 (bd, 1 H, J 7 Hz, NH), 7.16–7.37

(m, 25 H, 5 Ph). Anal. Calcd for $C_{46}H_{51}NO_7$: C, 75.69; H, 7.04; N, 1.92. Found: C, 75.40; H, 7.05; N, 1.70.

2,6-Anhydro-7-tert-butoxycarbonylamino-7-deoxy-7-C-phenyl-D-erythro-L-gulo-heptitol (21).—To a solution of 20 (220 mg, 0.301 mmol) in EtOAc (6 mL) and MeOH (6 mL) was added palladium-on-carbon (10%, 20 mg). After hydrogenolysis for 18 h, the mixture was filtered and concentrated under reduced pressure. Flash chromatography (9:1 CHCl₃-MeOH) yielded 21 (110 mg, 99%) as a colorless glassy solid; $[\alpha]_D$ +10.5° (c 1, MeOH); R_f 0.33 (9:1 CHCl₃-MeOH); ¹H NMR (250 MHz, CD₃OD): δ 1.31 [bs, 9 H, C(CH₃)₃], 3.42-3.77 (m, 6 H, H-1a,1b,2,3,4,5), 4.01 (dd, 1 H, $I_{5,6}$ 2.6, $I_{6,7}$ 7.3 Hz, H-6), 4.90 (d, 1 H, $I_{6,7}$ Hz, H-7), 7.12-7.33 (m, 5 H, Ph).

1,3,4,5-Tetra-O-acetyl-2,6-anhydro-7-tert-butoxycarbonylamino-7-deoxy-7-C-phenyl-p-erythro-L-gulo-heptitol (22).—To a solution of 21 (38 mg, 0.102 mmol) in dry pyridine (2 mL) was added Ac₂O (2 mL), and the mixture was left at room temperature overnight. When the reaction was complete as shown by TLC [R_f 0.5 (100:2 CHCl₃-MeOH)], the mixture was coevaporated with toluene (3 × 20 mL). Flash chromatography (100:2 CHCl₃-MeOH) yielded 22 (55 mg, quant) as a colorless crystalline solid; mp 209°C (with dec); $[\alpha]_D + 16^\circ$ (c 0.125, CHCl₃); ¹H NMR (250 MHz, CDCl₃): δ 1.36 [s, 9 H, C(CH₃)₃], 1.84, 2.04, 2.07, 2.09 (4 s, 12 H, 4 OAc), 3.89 (dd, 1 H, J_{gem} 11.7, $J_{1a,2}$ 3.8 Hz, H-1a), 3.97–4.08 (m, 1 H, H-2), 4.22-4.30 (m, 2 H, H-1b,5), 4.78-4.84 (m, 2 H, H-3,7), 5.02 (t, 1 H, J 9.3 Hz, H-6), 5.06 (dd, 1 H, $J_{3.4}$ 4.3, $J_{4.5}$ 3.1 Hz, H-4), 5.18 (m, 1 H, NH), 7.23–7.35 (m, 5 H, Ph). 2,6-Anhydro-1,3,4,5-tetra-O-benzyl-7-tert-butoxycarbonylamino-7-deoxy-7-Cphenyl-L-threo-L-gulo-heptitol (24).—Compound 23 (63 mg, 0.1 mmol) in dioxane (5 mL) was treated with (tBuOCO)₂O (71 mg) and Et₃N (0.5 mL) as described for 20. Flash chromatography $(7:1 \rightarrow 5:1 \text{ light petroleum-EtOAc})$ yielded 24 (66 mg, 90%) as a colorless oil; $[\alpha]_D + 27^\circ$ (c 1, CHCl₃); R_f 0.42 (4:1 light petroleum-EtOAc); ¹H NMR (250 MHz, CDCl₃): δ 1.31 [bs, 9 H, C(CH₃)₃], 3.46 (dd, 1 H, $J_{4.5}$ 5.9, $J_{5.6}$ 4.0 Hz, H-5), 3.63-3.76 (m, 3 H, H-1a,1b,3), 3.90 (t, 1 H, J 5.9 Hz, H-4), 3.90 (d, 1 H, J 11.4 Hz, CHHPh), 3.97 (m, 1 H, H-2), 4.13 (dd, 1 H, $J_{5.6}$ 4.0, $J_{6.7}$ 9.4 Hz, H-6), 4.23 (d, 1 H, J 11.4 Hz, CH H Ph), 4.45-4.70 (m, 6 H, 3 CH₂Ph), 4.90 (bs, 1 H, H-7), 5.25 (bd, 1 H, J 4.8 Hz, NH), 7.04-7.35 (m, 25 H, 5 Ph). Anal. Calcd for C₄₆H₅₁NO₇: C, 75.69; H, 7.04; N, 1.92. Found: C, 75.25; H, 7.07; N, 1.78. 2,6-Anhydro-7-tert-butoxycarbonylamino-7-deoxy-7-C-phenyl-L-threo-L-gulo-heptitol (25).—Compound 24 (210 mg, 0.288 mmol) in EtOAc (6 mL) and MeOH (6 mL) was treated with palladium-on-carbon (10%, 20 mg) as described for 21. Flash chromatography (9:1 CHCl₃-MeOH) of the residue yielded 25 (99 mg, 93%) as a colorless foam; $[\alpha]_D + 39.5^\circ$ (c 1, CHCl₃); R_f 0.26 (9:1 CHCl₃-MeOH); ¹H NMR (250 MHz, CD_3OD): δ 1.40 [bs, 9 H, $C(CH_3)_3$], 3.16–3.21 (m, 1 H, H-5), 3.58 (ddd, 1 H, $J_{2,3} = J_{3,4} = 4$, ${}^{4}J_{3,5}$ 0.7 Hz, H-3), 3.67 (dd, 1 H, J_{gem} 11.9, $J_{1a,2}$ 3.5 Hz, H-1a), 3.79 (t, 1 H, J 4 Hz, H-4), 3.97 (ddd, 1 H, $J_{1a,2} = J_{2,3} = 4$, $J_{1b,2}$ 8 Hz, H-2), 4.03 (dd, 1 H, $J_{5,6}$ 1.9, $J_{6,7}$ 10.2 Hz, H-6), 4.11 (dd, 1 H, J_{gem} 11.9, $J_{1\text{b},2}$ 8 Hz, H-1b), 4.90 (d, 1 H, J 10.2 Hz, H-7), 7.27–7.45 (m, 5 H, Ph).

1,3,4,5-Tetra-O-acetyl-2,6-anhydro-7-tert-butoxycarbonylamino-7-deoxy-7-C-phenyl-L-threo-L-gulo-heptitol (26).—Compound 25 (31 mg, 83.7 μ mol) was treated with dry pyridine (2 mL) and Ac₂O (2 mL) as described for 22. Flash chromatography (1:1 light petroleum–EtOAc) of the residue yielded 26 (45 mg, quant) as a colorless glassy foam; $[\alpha]_D + 39^\circ$ (c 1, CHCl₃); R_f 0.55 (1:1 light petroleum–EtOAc); ¹H NMR (250 MHz, CDCl₃): δ 1.34 [s, 9 H, C(CH₃)₃], 1.61, 1.99, 2.05, 2.11 (4 s, 12 H, 4 OAc), 3.96 (dd, 1 H, J_{gem} 12, $J_{\text{1a,2}}$ 2.5 Hz, H-1a), 4.13 (ddd, 1 H, $J_{\text{1a,2}}$ 2.5, $J_{\text{1b,2}} = J_{2,3} = 6.5$ Hz, H-2), 4.33 (dd, 1 H, $J_{3,4}$ 9.9, $J_{4,5}$ 3.9 Hz, H-4), 4.55 (dd, 1 H, J_{gem} 12, $J_{\text{1b,2}}$ 6.5 Hz, H-1b), 4.63 (dd, 1 H, $J_{4,5}$ 4, $J_{5,6}$ 7 Hz, H-5), 4.85 (t, 1 H, J_{4} 4 Hz, H-3), 4.85–4.98 (m, 1 H, H-7), 5.26–5.32 (m, 2 H, H-6, NH), 7.20–7.33 (m, 5 H, Ph).

2,6-Anhydro-1,3,4,5-tetra-O-benzyl-7-deoxy-7-(N-phenyltrifluoroacetamido)-Dglycero-L-gulo-heptitol (28).—To a solution of 27 (164 mg, 0.26 mmol) in dry CH_2Cl_2 (25 mL) was added trifluoroacetic anhydride (100 μ L, 0.708 mmol) with stirring at 0°C. Then, dry pyridine (100 μ L, 1.24 mmol) was added dropwise and the mixture was allowed to attain room temperature overnight. When TLC (4:1 light petroleum-EtOAc) showed the absence of starting material, the mixture was diluted with toluene and coevaporated $(2 \times 25 \text{ mL})$. Flash chromatography (6:1)light petroleum-EtOAc) of the residue yielded 28 (174 mg, 92%) as a colorless oil; $[\alpha]_{D}$ +24° (c 1, CHCl₃); R_f 0.48 (4:1 light petroleum-EtOAc); ¹H NMR (250 MHz, CDCl₃): δ 3.42 (dd, 1 H, J_{gem} 10.4, $J_{1a,2}$ 1.3 Hz, H-1a), 3.52-3.78 (m, 5 H, H-1b,2,3,4,5), 3.93-4.05 (m, 1 H, H-6), 4.29-4.40 (m, 2 H, H-7a,7b), 4.43-4.94 (m, 8 H, 4 C H_2 Ph), 7.09–7.40 (m, 25 H, 5 Ph); ¹³C NMR (62.5 MHz, CDCl₃): δ 46.84 (C-7), 68.25, 71.27, 72.04, 73.16, 73.45, 74.96, 75.45, 77.56, 78.77, 82.25 (10 C, 4 CH_2Ph , C-1,2,3,4,5,6), 116.31 (q, 1 C, ${}^2J_{CF}$ 288.4 Hz, CF₃), 127.58–129.21 (m, 25 C, o-, m-, p-Ph), 137.65, 137.86, 138.13, 138.54, 138.90 (5 C, ipso-Ph), 157.09 (q, 1 C, ${}^{3}J_{C.F}$ 35.7 Hz, COCF₃). Anal. Calcd for $C_{43}H_{43}F_{3}NO_{6}$: C, 71.06; H, 5.96; N, 1.93. Found: C, 71.13; H, 5.87; N, ca. 2.00.

2,6-Anhydro-7-deoxy-7-(N-phenyltrifluoroacetamido)-D-glycero-L-gulo-heptitol (29).—To a solution of 28 (106 mg, 0.146 mmol) in EtOAc (2.5 mL) and MeOH (2.5 mL) was added palladium-on-carbon (10%, 11 mg). After hydrogenolysis for 2 h, the mixture was filtered and concentrated under reduced pressure. Flash chromatography (9:1 CHCl₃-MeOH) of the residue yielded 29 (53.3 mg, quant) as a colorless foam; $[\alpha]_D$ +45.5° (c 1, CHCl₃); R_f 0.24 (9:1 CHCl₃-MeOH); ¹H NMR (250 MHz, CD₃OD): δ 3.41 (dd, 1 H, J 8.7, 9.2 Hz, H-3/4), 3.52–3.75 (m, 5 H, H-1a,1b,2,3/4,5), 3.81 (dd, 1 H, J_{6,7a} 2.8, J_{gem} 14.4 Hz, H-7a), 4.13 (ddd, 1 H, J_{5,6} 6.0, J_{6,7a} 2.8, J_{6,7b} 11.6 Hz, H-6), 4.63 (dd, 1 H, J_{6,7b} 11.6, J_{gem} 14.4 Hz, H-7b), 7.45–7.53 (m, 5 H, Ph).

1,3,4,5-Tetra-O-acetyl-2,6-anhydro-7-deoxy-7 (N-phenyltrifluoroacetamido)-D-glycero-L-gulo-heptitol (30).—Compound 29 (30 mg, 0.082 mmol) was treated with dry pyridine (2.5 mL) and Ac₂O (2.5 mL) as described for 22. Flash chromatography (2:1 light petroleum–EtOAc) of the residue yielded 30 (44 mg, quant), as a colorless oil; $[\alpha]_D$ +54° (c 1, CHCl₃); R_f 0.57 (1:1 light petroleum–EtOAc); ¹H

NMR (250 MHz, CDCl₃): δ 1.95, 1.98, 1.99, 2.04 (4 s, 12 H, 4 OAc), 3.53 (dd, 1 H, $J_{6,7a}$ 2.7, J_{gem} 14.4 Hz, H-7a), 3.92–4.02 (m, 2 H, H-1a, 2), 4.13 (dd, 1 H, J_{gem} 12.0, $J_{1b,2}$ 4.9 Hz, H-1b), 4.29 (ddd, 1 H, $J_{5,6}$ 6, $J_{6,7a}$ 2.7, $J_{6,7b}$ 11.5 Hz, H-6), 4.59 (dd, 1 H, $J_{6,7b}$ 11.5, J_{gem} 14.4 Hz, H-7b), 4.95 (t, 1 H, J 9 Hz, H-3), 5.11 (dd, 1 H, $J_{4,5}$ 9, $J_{5,6}$ 6 Hz, H-5), 5.28 (t, 1 H, J 9 Hz, H-4), 7.25–7.29 (m, 2 H, Ph), 7.38–7.43 (m, 3 H, Ph). Anal. Calcd for $C_{23}H_{26}F_3NO_{10}$: C, 51.78; H, 4.91; N, 2.63. Found: C, 51.12; H, 4.62; N, 2.45.

7-Ammonio-2,6-anhydro-7-deoxy-7-C-phenyl-D-erythro-L-gulo-heptitol trifluoroacetate (31).—To a solution of 21 (92 mg, 0.255 mmol) in CHCl₃ (5 mL) was added trifluoroacetic acid (1 mL), and the mixture was left at room temperature. When TLC (8:2 CHCl₃-MeOH) showed the absence of starting material (ca. 2.5 h), the mixture was diluted with CHCl₃ (15 mL), concentrated in vacuo, and coevaporated again with CHCl₃ (2 × 20 mL), to give 31 (98 mg, quant) as a colorless oil that crystallized on standing. For enzymic studies, 31 was subjected to preparative HPLC chromatography (9:1 $\rm H_2O + 0.1\%$ trifluoroacetic acid-MeCN; flow: 12 mL/min; t_R 8.9 min; 86 bar), concentrated in vacuo, and lyophilized; [α]_D + 38.5° (c 0.7, $\rm H_2O$); ¹H NMR (250 MHz, $\rm D_2O$): δ 3.16–3.36 (m, 4 H, H-1a,1b,2,3), 3.69 (t, 1 H, $\rm J$ 8.8 Hz, H-4), 3.79 (dd, 1 H, $\rm J$ _{4.5} 8.8, $\rm J$ _{5.6} 5.2 Hz, H-5), 4.34 (dd, 1 H, $\rm J$ _{5.6} 5.2, $\rm J$ _{6.7} 9.9 Hz, H-6), 4.73 (d, 1 H, $\rm J$ 9.9 Hz, H-7), 7.30–7.37 (m, 5 H, Ph). Anal. Calcd for $\rm C$ ₁₅ $\rm H$ ₂₀ $\rm F$ ₃NO₇ · 0.5H₂O: C, 45.92; H, 5.40; N, 3.57. Found: C, 45.95; H, 5.33; N, 3.71.

7-Ammonio-2,6-anhydro-7-deoxy-7-C-phenyl-L-threo-L-gulo-heptitol trifluoroacetate (32).—Compound 25 (85 mg, 0.23 mmol) in CHCl₃ (5 mL) was treated with trifluoroacetic acid (1 mL) as described for 31, to give 32 (88 mg, quant) as a colorless oil that crystallized on standing. For enzymic studies, 32 was subjected to preparative HPLC (9:1 H₂O + 0.05% trifluoroacetic acid–MeCN; flow: 9 mL/min; t_R 11.77 min; 59 bar), concentrated in vacuo, and lyophilized; [α]_D +26° (c 0.87, H₂O); ¹H NMR (250 MHz, D₂O): δ 3.18 (dd, 1 H, $J_{4,5}$ 4.1, $J_{5,6}$ 2.4 Hz, H-5), 3.49 (t, 1 H, J 4.8 Hz, H-3), 3.54–3.58 (m, 1 H, H-1a), 3.63 (t, 1 H, J 4.8 Hz, H-4), 3.80–3.90 (m, 2 H, H-1b,2), 4.20 (dd, 1 H, $J_{5,6}$ 2.4, $J_{6,7}$ 9.5 Hz, H-6), 4.47 (d, 1 H, J 9.5 Hz, H-7), 7.31 (bs, 5 H, Ph). Anal. Calcd for C₁₅H₂₀F₃NO₇·0.5H₂O: C, 45.92; H, 5.40; N, 3.57. Found: C, 45.82; H, 5.51; N, 3.60.

2,6-Anhydro-7-deoxy-7-phenylammonio-D-glycero-L-gulo-heptitol trifluoroacetate (33).—To a solution of 29 (67 mg, 0.183 mmol) in MeOH (5 mL) and H₂O (0.33 mL) was added Na₂CO₃ (100 mg) with stirring. After 4 h at room temperature, the mixture was filtered and the filter cake washed with MeOH (5 mL). The combined filtrates were neutralized with Amberlite IR-120 (H⁺) resin, filtered, and evaporated to dryness. Preparative HPLC (9:1 H₂O + 0.05% trifluoroacetic acid-MeCN; flow: 15 mL/min; t_R 8.32 min, λ_{max} 207.5 nm), concentration in vacuo, and lyophilisaton yielded 33 (68 mg, 97%); [α]_D +44° (c 0.5, MeOH); λ_{max}^{MeOH} 205.8, 245.0, 292.8 nm; ¹H NMR (250 MHz, D₂O): δ 3.19 (t, 1 H, J 9.2 Hz, H-3), 3.35 (t, 1 H, J 9.2 Hz, H-4), 3.39–3.51 (m, 1 H, H-2), 3.54–3.62 (m, 4 H, H-1a,1b,5,7a), 3.74 (dd, 1 H, $J_{6,7b}$ 11, J_{gem} 13.7 Hz, H-7b), 4.14 (ddd, 1 H, $J_{5,6}$ 6.1, $J_{6,7a}$ 2.6, $J_{6,7b}$ 11

Hz, H-6), 7.28–7.38 (m, 5 H, Ph). Anal. Calcd for $C_{15}H_{20}F_3NO_7 \cdot H_2O$: C, 44.78; H, 5.02; N, 3.49. Found: C, 44.54; H, 4.88; N, 3.88.

7-Amino-2,6-anhydro-7-deoxy-7-C-phenyl-D-erythro-L-gulo-heptitol (1a).—A solution of 31 (25 mg, 0.065 mmol) in water was chromatographed through a short column charged with Amberlite IRA-400 (OH $^-$) resin. Evaporation of the solvent yielded 1a (17.5 mg, quant) as a colorless oil; 1 H NMR (250 MHz, D $_2$ O): δ 3.17–3.33 (m, 4 H, H-1a,1b,2,3), 3.72–3.76 (m, 2 H, H-4,5), 4.00–4.05 (m, 1 H, H-6), 4.25 (d, 1 H, J 10.3 Hz, H-7), 7.19–7.29 (m, 5 H, Ph).

7-Amino-2,6-anhydro-7-deoxy-7-C-phenyl-L-threo-L-gulo-heptitol (2a).—Compound 32 (20 mg, 52.2 μ mol) was treated as described for 1a, to give 2a (14 mg, quant) as a colorless oil; 1 H NMR (250 MHz, D₂O): δ 3.28 (dd, 1 H, $J_{4,5}$ 6.4, $J_{5,6}$ 3.6 Hz, H-5), 3.38 (t, 1 H, J 6.5 Hz, H-3), 3.59–3.73 (m, 3 H, H-1a,2,4), 3.79 (dd, 1 H, J_{gem} 11.1, $J_{1\text{b},2}$ 6.6 Hz, H-1b), 3.94 (dd, 1 H, $J_{5,6}$ 3.6, $J_{6,7}$ 9.9 Hz, H-6), 4.08 (d, 1 H, J 9.9 Hz, H-7), 7.19–7.26 (m, 5 H, Ph).

2,6-Anhydro-7-deoxy-7-C-phenyl-D-glycero-L-gulo-heptitol (3a).—To a solution of 18 (35 mg, 82.8 μ mol) in dry MeOH (10 mL) was added sodium methoxide (0.15 mL of a 0.2 M solution in MeOH). After 2.5 h at room temperature, the mixture was neutralized with Amberlite IR-120 (H⁺) resin, filtered, and concentrated under reduced pressure. Flash chromatography (8:2 CHCl₃-MeOH) of the residue yielded 3a (21 mg, quant) as a colorless oil; $[\alpha]_D$ +133° (c 1, MeOH); R_f 0.14 (9:1 CHCl₃-MeOH); ¹H NMR (250 MHz, CD₃OD): δ 2.93-3.05 (m, 2 H, H-7a,7b), 3.34-3.42 (m, 1 H, H-2), 3.63-3.78 (m, 5 H, H-1a,1b,3,4,5), 4.12-4.19 (m, 1 H, H-6), 7.18-7.31 (m, 5 H, Ph).

2,6-Anhydro-7-deoxy-7-phenylamino-D-glycero-L-gulo-heptitol (4a).—Compound 33 (18.5 mg, 48.1 μ mol) was treated as described for 1a, to give 4a (13 mg, quant) as a colorless oil; R_f 0.3 (85:15 CHCl₃-MeOH); ¹H NMR (250 MHz, D₂O): δ 3.17–3.34 (m, 3 H, H-3,7a,7b), 3.37–3.44 (m, 1 H, H-2), 3.49 (t, 1 H, J 9.4 Hz, H-4), 3.51–3.68 (m, 3 H, H-1a,1b,5), 4.07 (ddd, 1 H, $J_{5,6} = J_{6,7a} = 5.1$, $J_{6,7b}$ 7.8 Hz, H-6), 6.70–6.74 (m, 3 H, Ph), 7.09–7.16 (m, 2 H, Ph).

Inhibition studies.—(a) Materials. Buffer substances were purchased from Fluka and used as received. α -D-Glucosidase (yeast, EC 3.2.1.20) and p-nitrophenyl α -D-glucopyranoside (PNPG) were obtained from Boehringer Mannheim.

- (b) Preparation of solutions. The buffer consisted of 0.01 M piperazine 1,4-bis(2-ethanesulfonic acid) (PIPES), 0.02 M NaOAc, and 0.1 mM EDTA³. The pH was adjusted to 6.5 with NaOH (10 M). PNPG was dissolved in the buffer solution for enzyme assays. 0.95 mg of α -D-glucosidase lyophilisate [ca. 8.4 U/mg (30°C, PNPG as substrate)] was dissolved in 1 mL of buffer solution and used for assays without further dilution. For each inhibitor, inhibitor concentrations ranging from 0 to 3 K_i , were generally used to determined the K_i value. At each inhibitor concentration, six substrate concentrations, spanning 0.4 K_M to 4 K_M , were used.
- (c) Procedure for enzyme assays. To a 1-mL disposable cuvette was added buffer solution (940 μ L), inhibitor solution (20 μ L), and PNPG-solution (20 μ L). The solution was thermally equilibrated at 30°C and the reaction was started by

addition of 20 μ L of α -D-glucosidase solution. Liberation of p-nitrophenol ($\epsilon_{ONP^-, pH 6.5}$ 3204.5 M⁻¹ cm⁻¹)²⁶ was monitored, using a Philips PU 8740 UV/VIS-spectrophotometer, for 60 s, and the initial hydrolysis rate was calculated.

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